



TRAP5 (human) ImmunoCapture Enzyme-Activity Assay Kit

Manufactured by BioVendor.

ALX-850-300-KI01

96 wells (~80 tests)

(Version 16: August 13, 2008)

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For laboratory use only. Not for human or diagnostic use.

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Use only the current version of Product Data Sheet enclosed with the kit!

1. INTENDED USE

The ALX-850-300 Human TRAP 5 Assay is an ImmunoCapture Enzyme-Activity Assay for the quantitative measurement of human TRAP 5 in serum and EDTA plasma.

»» Features

- **European Union: for in vitro diagnostic use.**
Rest of the world: for research use only!
- The total assay time is less than 4 hours.
- The kit measures TRAP 5 proteins exhibiting enzyme activity
- Assay format is 96 wells.
- Quality Controls are recombinant TRAP 5 protein based. No animal sera are used.
- Calibrators are recombinant protein based.
- Components of the kit are provided ready to use, concentrated or lyophilized.

2. STORAGE, EXPIRATION

Store the complete kit at 2-8°C. Under these conditions, the kit is stable until the expiration date (see label on the box).

For stability of opened reagents see Chapter 9.

3. INTRODUCTION

TRAP 5 (serum band 5 tartrate-resistant acid phosphatase, TRACP 5; EC 3.1.3.2) is a glycoprotein of 35-37 kDa. TRAP 5 belongs to the most abundant enzymes in osteoclasts. It is expressed in certain differentiated cells of the mononuclear phagocyte system, particularly osteoclasts and alveolar macrophages, where it takes an active part in bone resorption process.

High blood levels of TRAP 5 are usually associated with active bone remodelling. Increased serum levels are observed during normal bone growth among healthy children.

Elevated serum TRAP levels have been detected in diseases characterized by increased bone resorption; Paget's disease of the bone, hemodialysis, primary hyperparathyroidism, metastatic malignancies involving bone resorption, multiple myeloma and bilaterally ovariectomized

women. Post-menopausal women have higher levels of serum than post-menopausal women on estrogen replacement therapy.

Therefore specific determination of TRAP 5 activity can be essential for clinical assessment of bone metabolism.

Areas of investigation:

Bone and cartilage metabolism

4. TEST PRINCIPLE

In the ALX-850-300 Human TRAP 5 Assay, Calibrators, Quality Control and samples are incubated in microplate wells pre-coated with monoclonal anti-human TRAP 5 antibody. After a thorough wash, TRAP 5 bound to the antibody is allowed to react with the pNPP substrate at pH 5.5. The reaction is stopped by addition of hydroxide solution and absorbance of the resulting yellow colour product is measured spectrophotometrically at 405 nm. The absorbance is proportional to the enzymatic activity of TRAP 5. A standard curve is constructed by plotting absorbance values versus enzyme activities of recombinant TRAP 5 calibrators, and enzyme activity of unknown samples are determined (U/l) using this standard curve.

5. PRECAUTIONS

- **For professional use only.**
- Wear gloves and laboratory coats when handling immunodiagnostic materials.
- Do not drink, eat or smoke in the areas where immunodiagnostic materials are being handled.
- This kit contains components of human origin. These materials were found non-reactive for HBsAg, HCV antibody and for HIV 1/2 antigen and antibody. However, these materials should be handled as potentially infectious, as no test can guarantee the complete absence of infectious agents.
- Avoid contact with the corrosive alkaline Stop Solution and Substrate Solution, which contains pNPP. Wear gloves and eye and clothing protection when handling these reagents. Stop and/or Substrate Solutions may cause skin/eyes irritation. In case of contact with the Stop Solution and the Substrate Solution wash skin/eyes thoroughly with water and seek medical attention, when necessary.
- The materials must not be pipetted by mouth.

6. TECHNICAL HINTS

- Reagents with different lot numbers should not be mixed.
- Use thoroughly clean glassware.
- Use deionized (distilled) water, stored in clean containers.
- Avoid any contamination among samples and reagents. For this purpose, disposable tips should be used for each sample and reagent.
- Substrate Solution should remain colourless until added to the plate. Keep Substrate Solution protected from light.
- Stop Solution should remain colourless until added to the plate. The colour developed in the wells will turn from blue to yellow immediately after the addition of the Stop Solution. Wells that are green in colour indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.
- Dispose of consumable materials and unused contents in accordance with applicable national regulatory requirements.

7. REAGENT SUPPLIED

<i>Kit Components</i>	<i>State</i>	<i>Quantity</i>
Antibody Coated Microtiter Strips	ready to use	96 wells
Dissociating Solution	ready to use	7 ml
Calibrator (0.2-4 U/l)	lyophilized	3x 2 vials
Quality Control	lyophilized	2 vials
Dilution Buffer	ready to use	20 ml
Wash Solution Concentrate (10x)	concentrated	100 ml
Substrate Buffer		2x 20 ml
Substrate (pNPP) Tablets		4 pcs
Stop Solution (0.2 M NaOH)	ready to use	13 ml
Product Data Sheet + Certificate of Analysis		1 pc

8. MATERIAL REQUIRED BUT NOT SUPPLIED

- Deionized (distilled) water
- Test tubes for diluting samples
- Glassware (graduated cylinder and bottle) for Wash Solution (Dilution Buffer)
- Precision pipettes to deliver 50-200 μ l with disposable tips
- Multichannel pipette to deliver 100 μ l with disposable tips
- Absorbent material (e.g. paper towels) for blotting the microtiter plate after washing
- Vortex mixer
- Orbital microplate shaker capable of approximately 300 rpm
- Microplate washer (optional). [Manual washing is possible but not preferable.]
- Microplate reader with 405 ± 10 nm filter
- Software package facilitating data generation and analysis (optional)

9. PREPARATION OF REAGENTS

- All reagents need to be brought to room temperature prior to use.
- Always prepare only the appropriate quantity of reagents for your test.
- Do not use components after the expiration date marked on their label.

- Assay reagents supplied ready to use:

Antibody Coated Microtiter Strips

Stability and storage:

Return the unused strips to the provided aluminium zip-sealed bag with desiccant and seal carefully. Remaining Microtiter Strips are stable 3 months stored at 2-8°C and protected from the moisture.

Dissociating Solution

Dilution Buffer

Stop Solution

Stability and storage:

Opened reagents are stable 3 months when stored at 2-8°C.

- Assay reagents supplied concentrated or lyophilized:

Human TRAP 5 Calibrators

Reconstitute the lyophilized Calibrators just prior to the assay. Add **250 µl** of deionized (distilled) water to the vial containing lyophilized Calibrator. Let it dissolve at least 15 minutes and mix thoroughly.

Prepared Calibrators are ready to use, do not dilute them.

Stability and storage:

The reconstituted Calibrators have to be used immediately or to be stored frozen at -20°C for 3 months. Avoid repeated freeze/thaw cycles.

Quality Control

Reconstitute Quality Control (QC) just prior to the assay. Add **60 µl** of deionized (distilled) water to the vial containing lyophilized Quality Control. Let it dissolve at least 15 minutes and mix thoroughly.

Dilute reconstituted Quality Control 4x with Dilution Buffer, e.g. 60 µl of sample + 180 µl of Dilution Buffer for duplicates. **Mix well** (not to foam). Vortex is recommended.

Stability and storage:

The reconstituted Quality Control must be used immediately or to be stored frozen at -20°C for 3 months. Avoid repeated freeze/thaw cycles.

Do not store the diluted Quality Control.

Substrate Solution:

Add 1 Substrate Tablet to 10 ml of the Substrate Buffer, let it dissolve and mix the solution thoroughly. The solution should be prepared before use, 10-15 minutes is needed to dissolve the tablet

Stability and storage:

The Substrate Solution must be used immediately.

Wash Solution Concentrate (10x)

Dilute Wash Solution Concentrate (10x) ten-fold in distilled water to prepare a 1x working solution. Example: 100 ml of Wash Solution Concentrate (10x)+ 900 ml of distilled water for use of all 96-wells.

Stability and storage:

The diluted Wash Solution is stable 1 month when stored at 2-8°C. Opened Wash Solution Concentrate (10x) is stable 3 months when stored at 2-8°C.

10. PREPARATION OF SAMPLES

The kit measures TRAP 5 in serum or EDTA plasma.

Samples should be assayed immediately after collection or should be stored at -20°C. Mix thoroughly thawed samples just prior to the assay and avoid repeated freeze/thaw cycles, which may cause erroneous results. Avoid using hemolyzed or lipemic samples.

Dilute samples 4x with Dilution Buffer just prior to the assay, e.g. 60 µl of sample + 180 µl of Dilution Buffer for duplicates. **Mix well** (not to foam). Vortex is recommended.

Stability and storage:

Samples should be stored for 1 day at + 4°C, for 1 month at -20°C, or preferably at -80°C for six months. Avoid repeated freeze/ thaw cycles.

Do not store the diluted samples.

Note: It is recommended to use a precision pipette and a careful technique to perform the dilution in order to get precise results.

11. ASSAY PROCEDURE

1. Pipet **100 µl** of each Calibrators, diluted Quality Control, Dilution Buffer (=Blank) and diluted samples, preferably in duplicates, into the appropriate wells.
2. Pipet **50 µl** of Dissociating Solution into all the used wells.
3. Incubate the plate at room temperature (ca. 25°C) for **2 hours**, shaking at ca. 300 rpm on an orbital microplate shaker.
4. Wash the wells **3-times** with Wash Solution (0.35 ml per well). After final wash, invert and tap the plate strongly against paper towel.
5. Add **100 µl** of Substrate Solution into each well.
6. Incubate the plate in an incubator at 37°C for 1.5 hour, no shaking !
7. Stop the enzyme reaction by adding **100 µl** of Stop Solution.
8. Determine the absorbance by reading the plate at 405 nm (optionally, to measure in dual wavelength mode 620-650 nm filter can be used to measure the reference absorbance).

Warning

THE SUBSTRATE SOLUTION IS COLOURLESS DURING INCUBATION UNTILL ITS PH IS CHANGED BY ADDING THE STOP SOLUTION !!!

	strip 1+2	strip 3+4	strip 5+6	strip 7+8	strip 9+10	strip 11+12
A	Calibrator 4	Sample 4	Sample 12	Sample 20	Sample 28	Sample 36
B	Calibrator 2	Sample 5	Sample 13	Sample 21	Sample 29	Sample 37
C	Calibrator 0.2	Sample 6	Sample 14	Sample 22	Sample 30	Sample 38
D	Blank	Sample 7	Sample 15	Sample 23	Sample 31	Sample 39
E	Quality Control	Sample 8	Sample 16	Sample 24	Sample 32	Sample 40
F	Sample 1	Sample 9	Sample 17	Sample 25	Sample 33	Sample 41
G	Sample 2	Sample 10	Sample 18	Sample 26	Sample 34	Sample 42
H	Sample 3	Sample 11	Sample 19	Sample 27	Sample 35	Sample 43

Figure 1: Example of a work sheet.

12. CALCULATIONS

Most microplate readers perform automatic calculations of analyte concentration. The standard curve is constructed by plotting the mean absorbance at 405 nm (Y) of Calibrators against the known enzyme activity (X) of Calibrators, using linear regression function. Results are reported as enzyme activity of TRAP 5 (U/l) in samples.

The measured enzyme activity of samples and Quality Control calculated from the standard curve must be multiplied by their respective dilution factor, because samples and Quality Control have been diluted prior to the assay (e.g. 2.5 U/l (from standard curve) x 4 (dilution factor) = 10 U/l).

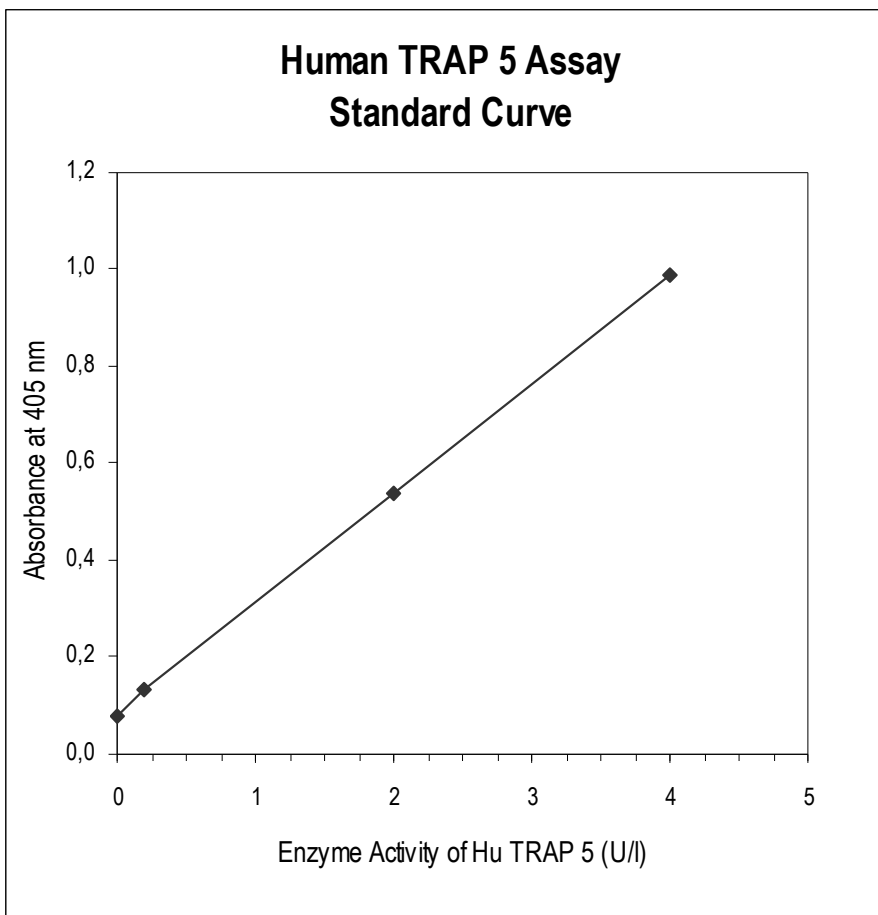


Figure 2: Typical Standard Curve for Human TRAP 5 Assay.

13. PERFORMANCE CHARACTERISTICS

➤➤ Typical analytical data of ALX-850-300 Human TRAP 5 Assay are presented in this chapter.

- **Sensitivity**

Limit of Detection (LOD) (defined as concentration of analyte giving absorbance higher than mean absorbance of blank* plus three standard deviations of the absorbance of blank: $A_{\text{blank}} + 3 \times \text{SD}_{\text{blank}}$) is calculated from the real TRAP 5 values in wells and is 0.01 U/l.

*Dilution Buffer is pipetted into blank wells.

- **Limit of assay**

Results exceeding TRAP 5 level of 16 U/l should be repeated with more diluted samples. Dilution factor needs to be taken into consideration in calculating the TRAP 5 concentration.

- **Specificity**

The antibodies used in this Assay are specific for human TRAP 5.

Sera of several mammalian species were measured in the assay. See results below.

<i>Mammalian serum sample</i>	<i>Observed crossreactivity</i>
Bovine	no
Cat	no
Dog	yes
Goat	no
Hamster	no
Horse	no
Monkey	yes
Mouse	no
Pig	no
Rabbit	yes
Rat	no
Sheep	no

- Precision**

Intra-assay (Within-Run) (n=8)

<i>Sample</i>	<i>Mean (U/l)</i>	<i>SD (U/l)</i>	<i>CV (%)</i>
1	1.88	0.00	0.8
2	2.20	0.01	2.4

Inter assay (Run-to-Run) (n=8)

<i>Sample</i>	<i>Mean (U/l)</i>	<i>SD (U/l)</i>	<i>CV (%)</i>
1	2.10	0.13	6.4
2	2.97	0.23	7.6

- Spiking Recovery**

Serum samples were spiked with different amounts of human TRAP 5 and assayed.

<i>Sample</i>	<i>Observed (U/l)</i>	<i>Expected (U/l)</i>	<i>Recovery O/E (%)</i>
1	2.67	-	-
	2.80	2.87	98
	4.47	4.67	96
	6.26	6.67	94
2	3.26	-	-
	3.17	3.46	92
	4.81	5.26	91
	6.76	7.26	93

- Linearity**

Serum samples were serially diluted with Dilution Buffer (4x) and assayed.

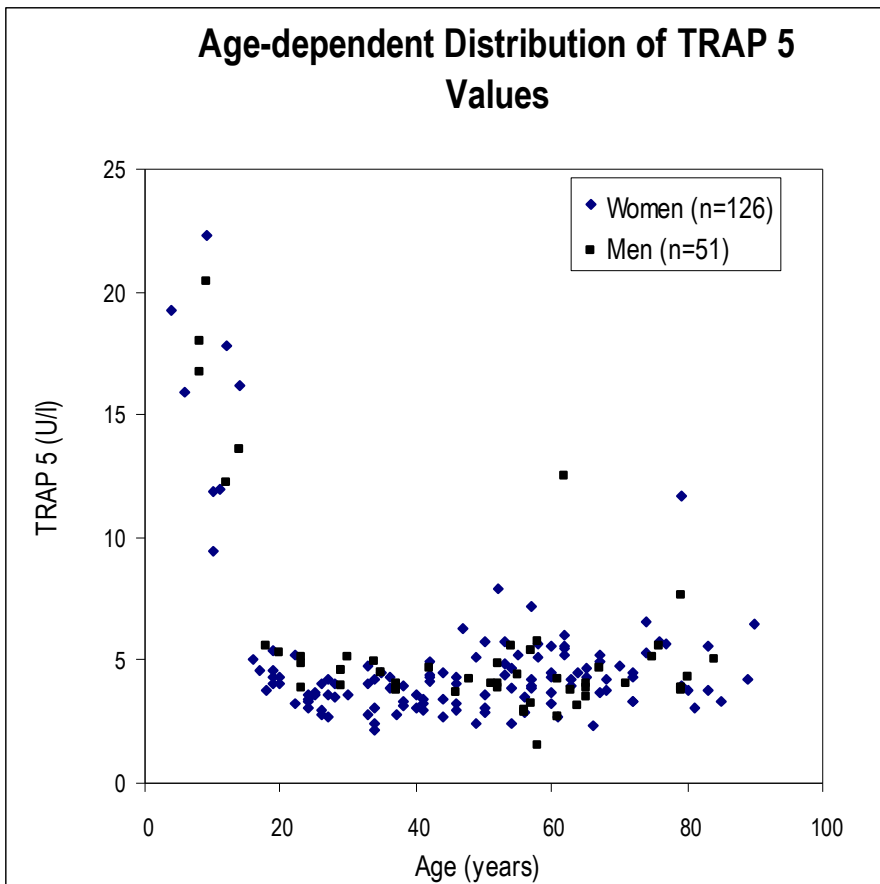
<i>Sample</i>	<i>Dilution</i>	<i>Observed (U/l)</i>	<i>Expected (U/l)</i>	<i>Recovery O/E (%)</i>
1	-	7.83	-	-
	2x	4.31	3.92	110
	4x	2.05	1.96	105
	8x	1.05	0.98	107
2	-	5.27	-	-
	2x	2.63	2.64	100
	4x	1.37	1.32	104
	8x	0.71	0.66	108

- **Reference Ranges**

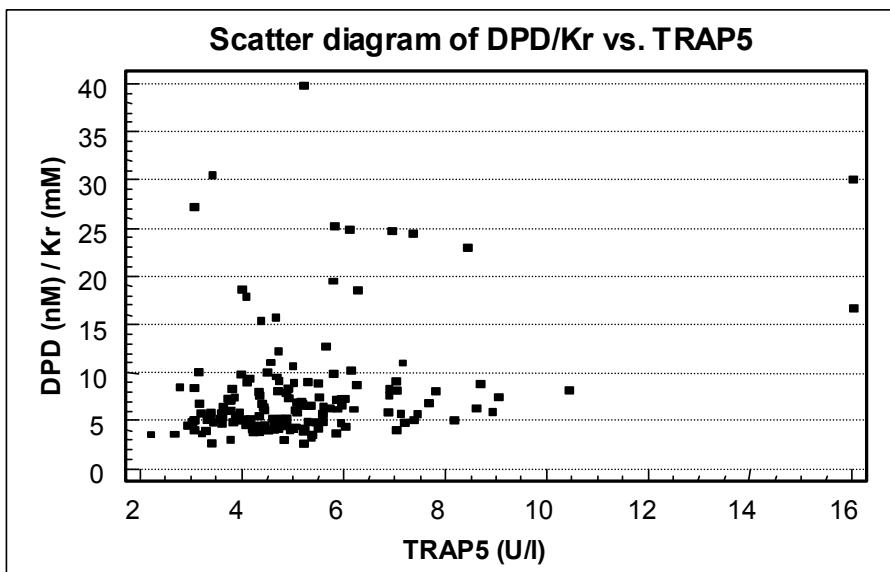
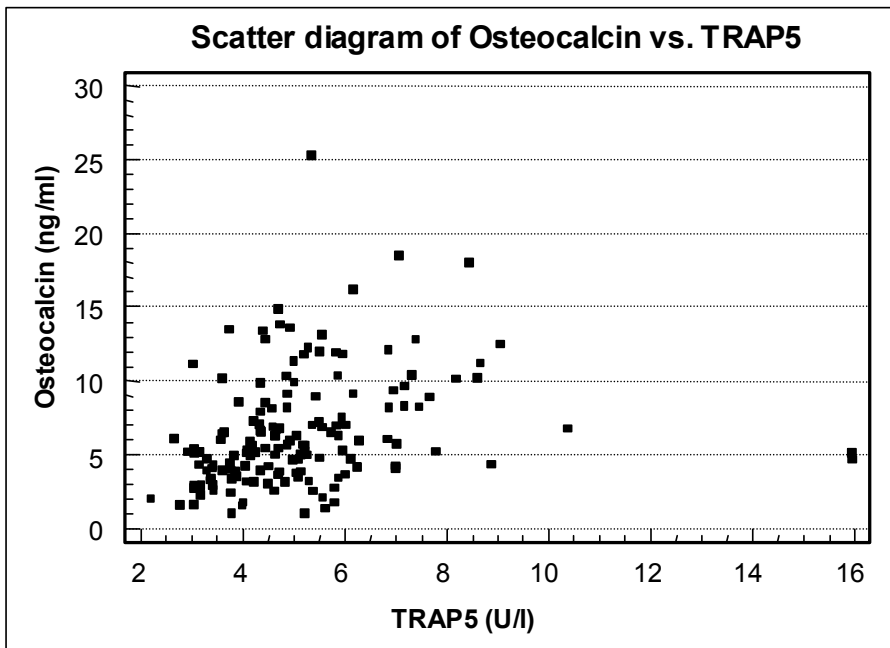
The data quoted in these instructions should be used for guidance only. It is recommended that each laboratory include its own panel of control sample in the assay. Each laboratory should establish its own normal and pathological references ranges for TRAP 5 levels with the assay.

a) The following results were obtained when 177 random samples were analysed with ALX-850-300 Human TRAP 5 Assay.

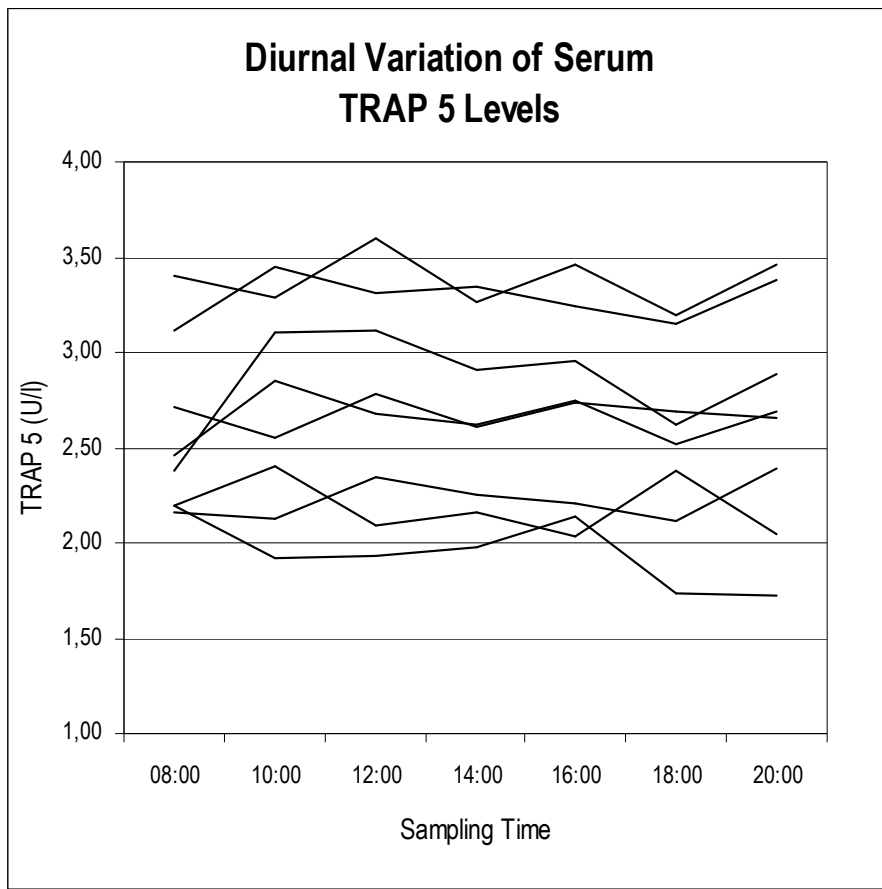
<i>Gender</i>	<i>Age (years)</i>	<i>n</i>	<i>Mean (U/l)</i>	<i>SD (U/l)</i>	<i>Min (U/l)</i>	<i>Max (U/l)</i>
Men	8-14	5	16.18	3.29	12.25	20.38
	18-48	16	4.54	0.58	3.70	5.61
	51-84	30	4.47	1.90	1.56	12.50
Women	4-14	8	15.58	4.28	9.40	22.26
	16-50	63	3.79	0.85	2.12	6.32
	52-90	55	4.68	1.49	2.38	11.69



b) TRAP 5 levels were determined in 489 patients with osteopathy and were compared with those of osteocalcin and DPD/Kr.



c) Diurnal variation of TRAP 5 levels in serum was determined in 8 patients in the course of 12 hours.

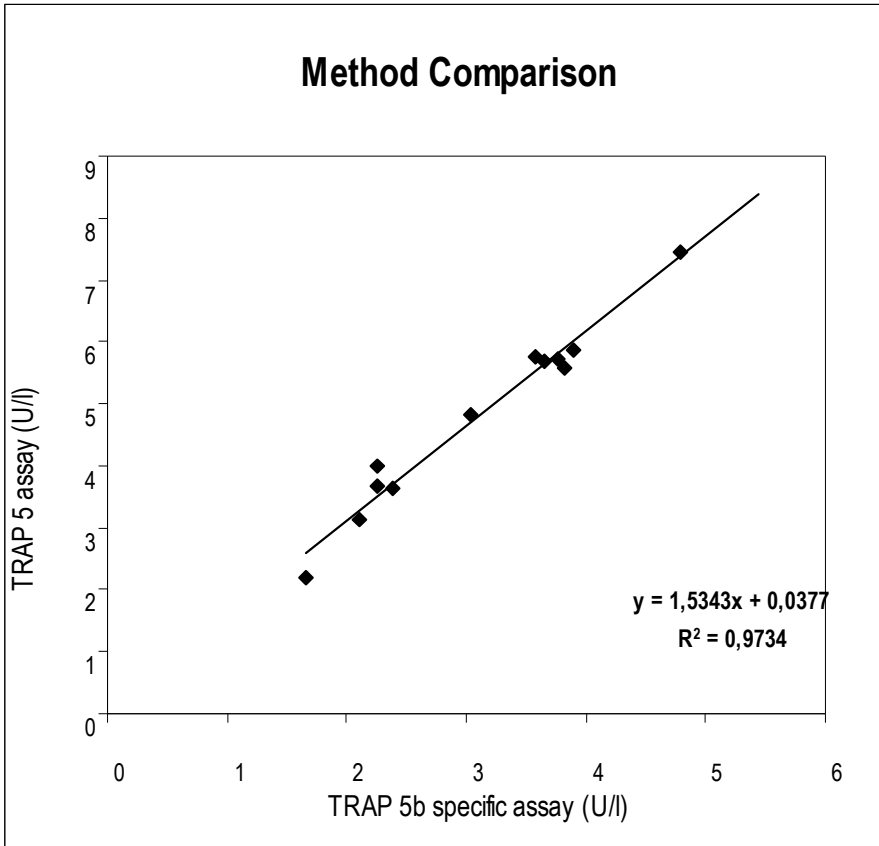


14. DEFINITION OF THE CALIBRATOR

The recombinant protein is used as the Calibrator in this assay. Recombinant human TRAP is expressed in Sf 9 cell.

15. METHOD COMPARISON

The ALX-850-300 Human TRAP 5 Assay was compared to a commercial assay specific for TRAP 5b isoform. Linear regression analysis of the results yielded the following results:



16. TROUBLESHOOTING AND FAQs

➤➤ **Weak signal in all wells**

Possible explanations:

- Omission of a reagent or a step
- Improper preparation or storage of a reagent
- Assay performed before reagents were allowed to come to room temperature
- Improper wavelength when reading absorbance

➤➤ **High signal and background in all wells**

Possible explanations:

- Improper or inadequate washing
- Overdeveloping; incubation time with Substrate Solution should be decreased before addition of Stop Solution
- Incubation temperature over 30°C

➤➤ **High coefficient of variation (CV)**

Possible explanation:










- Improper or inadequate washing
- Improper mixing Calibrators, Quality Controls or samples

17. REFERENCES

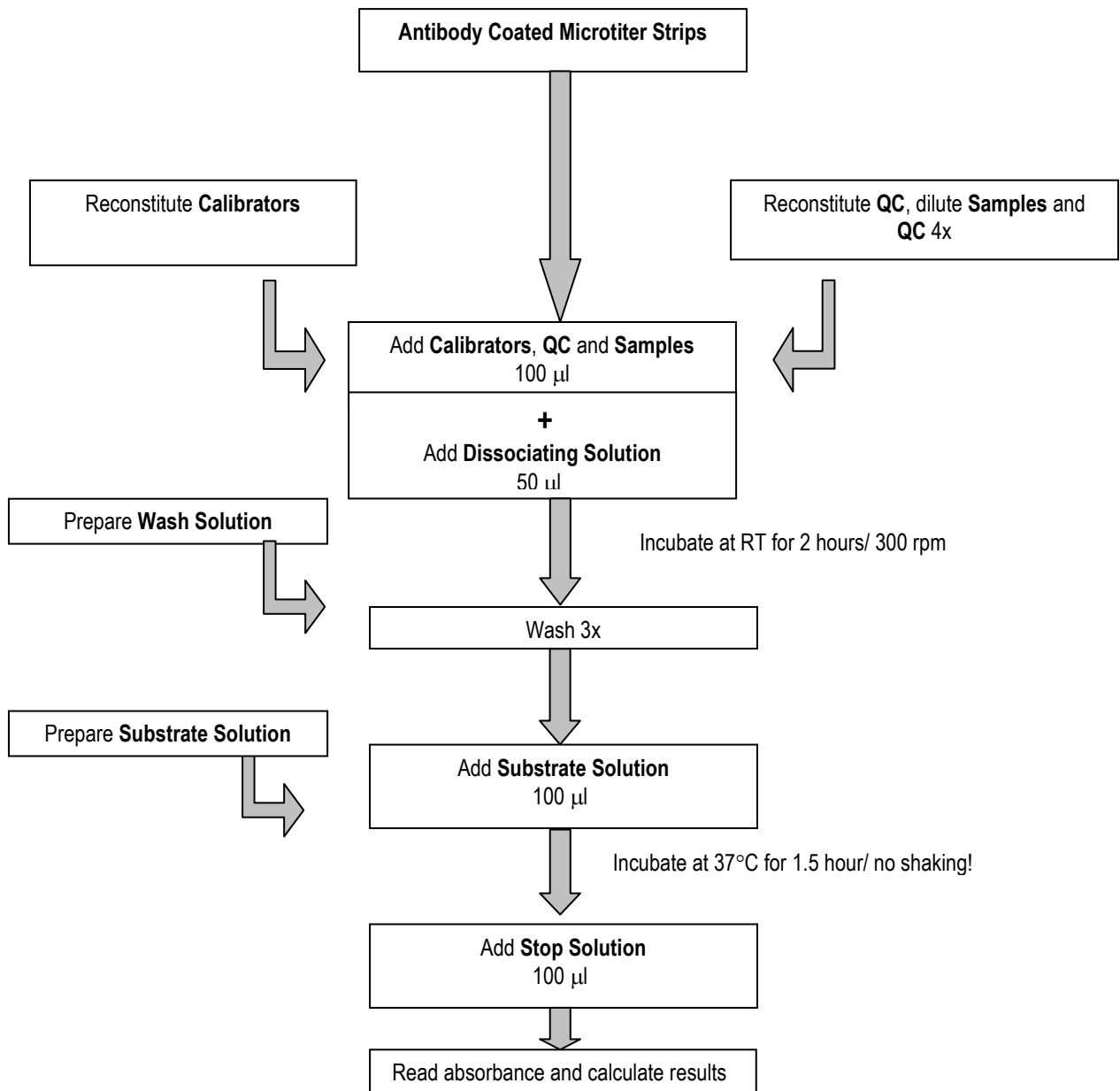
»» References to Human TRAP 5 Assay:

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- Nakanishi M, Yoh K, Miura T, Ohasi T, Rai SK, Uchida K. Development of a kinetic assay for band 5b tartrate-resistant acid phosphatase activity in serum. *Clin Chem.* 2000, 46, 469-473.
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- Igarashi Y, Lee M, Matsuzaki S. Acid phosphatases as markers of bone metabolism. *J Chromatogr B.* 2002, 781, 345-358.
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18. EXPLANATION OF SYMBOLS

	<p>Catalogue number</p>
	<p>Content</p>
	<p>Lot number</p>
	<p>See instructions for use</p>
	<p>Biological hazard</p>
	<p>Expiry date</p>
	<p>Storage conditions</p>
	<p>Identification of packaging materials</p>
	<p>In vitro diagnostic medical device</p>

Assay Procedure Summary



12								
11								
10								
9								
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6								
5								
4								
3								
2								
1								
	A	B	C	D	E	F	G	H

NOTES



