



Cystatin C (rat) ELISA Kit

Manufactured by BioVendor.

ALX-850-330-KI01

96 wells (~80 tests)

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For laboratory use only. Not for human or diagnostic use.

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Use only the current version of Product Data Sheet enclosed with the kit!

1. INTENDED USE

The ALX-850-330 Rat Cystatin C ELISA is a sandwich enzyme immunoassay for the quantitative measurement of rat Cystatin C.

»» Features

- **It is intended for research use only.**
- The total assay time is less than 3.5 hours.
- The kit measures total Cystatin C in rat serum.
- Assay format is 96 wells.
- Quality Controls are animal serum based. No human sera are used.
- Standard is recombinant protein based.
- Components of the kit are provided ready to use, concentrated or lyophilized.

2. STORAGE, EXPIRATION

Store the complete kit at 2-8°C. Under these conditions, the kit is stable until the expiration date (see label on the box).

For stability of opened reagents see Chapter 9.

3. INTRODUCTION

Cystatin C is a mammalian cysteine protease inhibitor, synthesized in various levels by different cell-types and appears in most body fluids.

Cystatin C has nothing to do with the statin drugs that are used to lower cholesterol. It is also known as cystatin 3 and CST3.

Cystatins belong to a superfamily of cysteine proteases inhibitors such as papain and Cathepsins B, H, K, L, and S. They have been found in both plants and animals. Cystatin C, with molecular weight of 13 260 Da and composed of 120 amino acids, lacks carbohydrate and contains two disulfide bridges located near the carboxyl terminus.

Cystatin C was identified, quantitated, and localized in mouse, rat, and human retinas. In the normal adult rat retina Cystatin C is present at high concentrations as it is throughout its postnatal development. Its concentration increases to a peak at the time when rat pups open their eyes and remains at a high level. It is mainly localized to the pigment epithelium, but also to some few neurons of varying types in the inner retina. Cystatin C is similarly expressed in normal mouse and human retinas.

Cystatin C is in the anterior segment of normal rat and mouse eyes. Cysteine proteases play an important role in protein degradation (e.g. of photoreceptor outer segments in the retinal pigment epithelium) and the balance between these proteases and their specific inhibitors is therefore of great interest.

Cystatin C level is increased in patients with malignant diseases, rheumatic diseases and related to the insufficiency of renal function. This protein appears to be a better marker than creatine. It may be especially useful in those cases where the creatinine measurement is not appropriate: for instance in liver cirrhosis, an obese, in malnourished or in patients with reduced muscle mass too Cystatin C measurement may also be useful in the early detection of kidney disease when other parameters might still be normal, especially in the elderly. In addition to kidney dysfunction, it has been associated with an increased risk of cardiovascular disease and heart failure in older adults. Researches are exploring other uses of Cystatin C, and the reasons for doctors ordering they may evolve over time.

Low levels of Cystatin C indicate cause the breakdown of the elastic laminae and, subsequently, the atherosclerosis and abdominal aortic aneurysm. The blood level of Cystatin C predicts survival after one type of heart attack. On the other hand, a high level of Cystatin C in the blood after a heart attack is an ominous sign because it reflects the failure of kidney to clear Cystatin C from the blood into the urine.

A mutation of the Cystatin C gene is responsible for a type of amyloidosis in which deposits in the brain result in premature strokes, intracranial hemorrhage, and dementia. This disease is called amyloidosis VI or cerebroarterial amyloidosis. It is inherited in an autosomal dominant manner.

Areas of investigation:

Kidney disease

Oncology

Rheumatism

4. TEST PRINCIPLE

In the ALX-850-330 Rat Cystatin C ELISA, Standards, Quality Controls and samples are incubated in microplate wells pre-coated with polyclonal anti-rat Cystatin C antibody. After 60 minutes incubation and washing, biotin-labelled polyclonal anti- mouse Cystatin antibody is added and incubated for 60 minutes with captured cystatin C. After another washing, streptavidin-HRP conjugate is added. After 30 minutes incubation and the last washing step, the remaining conjugate is allowed to react with the substrate solution (TMB). The reaction is stopped by addition of acidic solution and absorbance of the resulting yellow product is measured spectrophotometrically at 450 nm. The absorbance is proportional to the concentration of Cystatin C. A standard curve is constructed by plotting absorbance values against concentrations of Standards, and concentrations of unknown samples are determined using this standard curve.

5. PRECAUTIONS

- **For professional use only.**
- Wear gloves and laboratory coats when handling immunodiagnostic materials.
- Do not drink, eat or smoke in the areas where immunodiagnostic materials are being handled.
- This kit contains components of mouse origin. However, these materials should be handled as potentially infectious, as no test can guarantee the complete absence of infectious agents.
- Avoid contact with the acidic Stop Solution and Substrate Solution, which contains hydrogen peroxide and tetramethylbenzidine (TMB). Wear gloves and eye and clothing protection when handling these reagents. Stop and/or Substrate Solutions may cause skin/eyes irritation. In case of contact with the Stop Solution and the Substrate Solution wash skin/eyes thoroughly with water and seek medical attention, when necessary.
- The materials must not be pipetted by mouth.

6. TECHNICAL HINTS

- Reagents with different lot numbers should not be mixed.
- Use thoroughly clean glassware.
- Use deionized (distilled) water, stored in clean containers.
- Avoid any contamination among samples and reagents. For this purpose, disposable tips should be used for each sample and reagent.
- Substrate Solution should remain colourless until added to the plate. Keep Substrate Solution protected from light.
- Stop Solution should remain colourless until added to the plate. The colour developed in the wells will turn from blue to yellow immediately after the addition of the Stop Solution. Wells that are green in colour indicate that the Stop Solution has not mixed thoroughly with the Substrate Solution.
- Dispose of consumable materials and unused contents in accordance with applicable national regulatory requirements.

7. REAGENT SUPPLIED

<i>Kit Components</i>	<i>State</i>	<i>Quantity</i>
Antibody Coated Microtiter Strips	ready to use	96 wells
Biotin Labelled Antibody	ready to use	13 ml
Streptavidin-HRP Conjugate	ready to use	13 ml
Master Standard	lyophilized	2 vials
Quality Control High	lyophilized	2 vials
Quality Control Low	lyophilized	2 vials
Dilution Buffer	ready to use	2 x 20 ml
Wash Solution Concentrate (10x)	concentrated	100 ml
Substrate Solution	ready to use	13 ml
Stop Solution	ready to use	13 ml
Product Data Sheet + Certificate of Analysis		1 pc

8. MATERIAL REQUIRED BUT NOT SUPPLIED

- Deionized (distilled) water
- Test tubes for diluting samples
- Glassware (graduated cylinder and bottle) for Wash Solution (Dilution Buffer)
- Precision pipettes to deliver 5-1000 μl with disposable tips
- Multichannel pipette to deliver 100 μl with disposable tips
- Absorbent material (e.g. paper towels) for blotting the microtiter plate after washing
- Vortex mixer
- Orbital microplate shaker capable of approximately 300 rpm
- Microplate washer (optional). [Manual washing is possible but not preferable.]
- Microplate reader with 450 ± 10 nm filter
- Software package facilitating data generation and analysis (optional)

9. PREPARATION OF REAGENTS

- All reagents need to be brought to room temperature prior to use.
- Always prepare only the appropriate quantity of reagents for your test.
- Do not use components after the expiration date marked on their label.

- Assay reagents supplied ready to use:

Antibody Coated Microtiter Strips

Stability and storage:

Return the unused strips to the provided aluminium zip-sealed bag with desiccant and seal carefully. Remaining Microtiter Strips are stable 3 months stored at 2-8°C and protected from the moisture.

Biotin Labelled Antibody

Streptavidin-HRP Conjugate

Dilution Buffer

Substrate Solution

Stop Solution

Stability and storage:

Opened reagents are stable 3 months when stored at 2-8°C.

- Assay reagents supplied concentrated or lyophilized:

Rat Cystatin C Master Standard

Refer to Certificate of Analysis for actual volume of Dilution Buffer needed for reconstitution of standard!!!

Reconstitute the lyophilized Master Standard with Dilution Buffer just prior to the assay. Let it dissolve at least 15 minutes with occasional gentle shaking (not to foam). The resulting concentration of the rat Cystatin C in the stock solution is **50 ng/ml**.

Prepare set of standards using Dilution Buffer as follows:

Volume of Standard	Standard Diluent	Concentration
Stock	-	50 ng/ml
250 µl of stock	250 µl	25 ng/ml
250 µl of 25 ng/ml	250 µl	12.5 ng/ml
250 µl of 12.5 ng/ml	250 µl	6.25 ng/ml
250 µl of 6.25 ng/ml	250 µl	3.13 ng/ml
250 µl of 3.125 ng/ml	250 µl	1.56 ng/ml
250 µl of 1.56 ng/ml	250 µl	0.78 ng/ml

Prepared Standards are ready to use, do not dilute them.

Stability and storage:

Standard stock solution (10 ng/ml) should be aliquoted and frozen at -20°C for 3 months.

Avoid repeated freeze/thaw cycles.

Do not store the diluted Standard Solutions.

Quality Controls High, Low

IMPORTANT: Refer to the Certificate of Analysis for actual volume of Dilution Buffer needed for reconstitution and for actual Quality Controls concentrations.

Reconstitute each Quality Control (High and Low) with Dilution Buffer just prior to the assay. Let it dissolve at least 15 minutes with occasional gentle shaking (not to foam).

Dilute reconstituted Quality Controls 100x with Dilution Buffer, e.g. 3 µl of Quality Control + 297 µl of Dilution Buffer for duplicates. Mix well (not to foam). Vortex is recommended.

Stability and storage:

The reconstituted Quality Controls must be used immediately.

Avoid repeated freeze/thaw cycles.

Do not store the diluted Quality Controls.

Wash Solution

Dilute Wash Solution Concentrate (10x) ten-fold in 900 ml of distilled water to prepare a 1x working solution.

Example: 100 ml of Wash Solution Concentrate (10x) + 900 ml of distilled water for use of all 96-wells.

Stability and storage:

The diluted Wash Solution is stable 1 month when stored at 2-8°C. Opened Wash Solution Concentrate (10x) is stable 3 months when stored at 2-8°C.

10. PREPARATION OF SAMPLES

The kit measures Cystatin C in rat serum.

Samples should be assayed immediately after collection or should be stored at -20°C. Mix thoroughly thawed samples just prior to the assay and avoid repeated freeze/thaw cycles, which may cause erroneous results. Avoid using hemolyzed or lipemic samples.

Dilute samples just prior to the assay 100x with Dilution Buffer follows:

Add 3 µl of sample into 297 µl of Dilution Buffer for duplicates. Mix well (not to foam). Vortex is recommended.

Stability and storage:

Samples should be stored at -20°, or preferably at -70°C for long-term storage. Avoid repeated freeze/ thaw cycles.

Do not store the diluted samples.

See Chapter 13 for stability of serum if stored at 2-8°C, effect of freezing/thawing on the concentration of Cystatin C.

Note: It is recommended to use a precision pipette and a careful technique to perform the dilution in order to get precise results.

11. ASSAY PROCEDURE

1. Pipet **100 µl** of diluted Standards, Quality Controls, Dilution Buffer (=Blank) and samples, preferably in duplicates, into the appropriate wells. See Figure 1 for example of work sheet.
2. Incubate the plate at room temperature (ca. 25°C) for **1 hour**, shaking at ca. 300 rpm on an orbital microplate shaker.
3. Wash the wells **3-times** with Wash Solution (0.35 ml per well). After final wash, invert and tap the plate strongly against paper towel.
4. Add **100 µl** of Biotin Labeled Antibody into each well.
5. Incubate the plate at room temperature (ca. 25°C) for **1 hour**, shaking at ca. 300 rpm on an orbital microplate shaker.
6. Wash the wells **3-times** with Wash Solution (0.35 ml per well). After final wash, invert and tap the plate strongly against paper towel.
7. Add **100 µl** of Streptavidin-HRP Conjugate into each well.
8. Incubate the plate at room temperature (ca. 25°C) for **30 minutes**, shaking at ca. 300 rpm on an orbital microplate shaker.
9. Wash the wells **3-times** with Wash Solution (0.35 ml per well). After final wash, invert and tap the plate strongly against paper towel.
10. Add **100 µl** of Substrate Solution into each well. Avoid exposing the microtiter plate to direct sunlight. Covering the plate with e.g. aluminium foil is recommended.
11. Incubate the plate for **10 minutes** at room temperature. The incubation time may be extended [up to 20 minutes] if the reaction temperature is below than 20°C. Do not shake the plate during the incubation.
12. Stop the colour development by adding **100 µl** of Stop Solution.
13. Determine the absorbance by reading the plate at 450 nm. The absorbance should be read within 5- 15 minutes following step 12.

Note: If the microplate reader is not capable of reading absorbance greater than the absorbance of the highest standard, perform a second reading at 405 nm. A new standard curve, constructed using the values measured at 405 nm, is used to determine Cystatin C concentration of off-scale samples. The readings at 405 nm should not replace the on-scale readings at 450 nm.

	strip 1+2	strip 3+4	strip 5+6	strip 7+8	strip 9+10	strip 11+12
A	Standard 50	QC Low	Sample 7	Sample 15	Sample 23	Sample 31
B	Standard 25	QC High	Sample 8	Sample 16	Sample 24	Sample 32
C	Standard 12.5	Sample 1	Sample 9	Sample 17	Sample 25	Sample 33
D	Standard 6.25	Sample 2	Sample 10	Sample 18	Sample 26	Sample 34
E	Standard 3.13	Sample 3	Sample 11	Sample 19	Sample 27	Sample 35
F	Standard 1.56	Sample 4	Sample 12	Sample 20	Sample 28	Sample 36
G	Standard 0.78	Sample 5	Sample 13	Sample 21	Sample 29	Sample 37
H	Blank	Sample 6	Sample 14	Sample 22	Sample 30	Sample 38

Figure 1: Example of a work sheet.

12. CALCULATIONS

Most microplate readers perform automatic calculations of analyte concentration. The Standard curve is constructed by plotting the mean absorbance at 450 nm (Y) of Standards against log of the known concentration (X) of Standards, using the four-parameter algorithm. Results are reported as concentration of cys C ng/ml in samples.

Alternatively, the logit log function can be used to linearize the standard curve (i.e. logit of the mean absorbance (Y) is plotted against log of the known concentration (X) of Standards.

The measured concentration of samples and Quality Controls calculated from the standard curve must be multiplied by their respective dilution factor, because samples and Quality Controls have been diluted prior to the assay. e.g. 10 ng/ml (from standard curve) x 100 (dilution factor) = 1000 ng/ml.

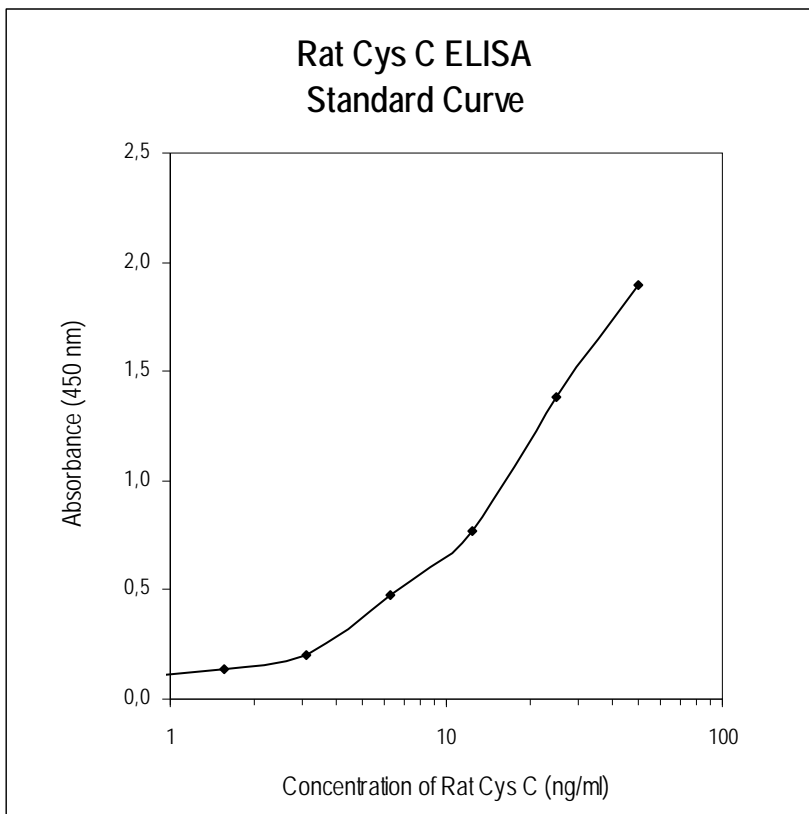


Figure 2: Typical Standard Curve for Rat Cystatin C ELISA

13. PERFORMANCE CHARACTERISTICS

➤➤ Typical analytical data of ALX850-330 Rat Cystatin C ELISA are presented in this chapter.

- **Sensitivity**

Limit of Detection (LOD) (defined as concentration of analyte giving absorbance higher than mean absorbance of blank* plus three standard deviations of the absorbance of blank: $A_{\text{blank}} + 3 \times \text{SD}_{\text{blank}}$) is calculated from the real Cystatin C values in wells and is 0.06 ng/ml.

*Dilution Buffer is pipetted into blank wells.

- **Limit of assay**

Results exceeding cys C level of 50 ng/ml should be repeated with more diluted samples. Dilution factor needs to be taken into consideration in calculating the Cystatin C concentration.

- **Specificity**

The antibodies in Rat Cystatin C ELISA kit are specific for rat Cystatin C. Approximately 83.4% cross-reactivity with recombinant mouse Cystatin C.

Sera of several mammalian species were measured in the assay. See results below.

<i>Mammalian serum sample</i>	<i>Observed crossreactivity</i>
Dog	no
Goat	no
Hamster	yes
Horse	no
Human	no
Monkey	no
Pig	no
Rabbit	no
Mouse	yes
Sheep	no

- **Precision**

Intra-assay (Within-Run) (n=8)

<i>Sample</i>	<i>Mean (ng/ml)</i>	<i>SD (ng/ml)</i>	<i>CV (%)</i>
1	590	0.27	4.7
2	235	0.16	6.6

Inter assay (Run-to-Run) (n=5)

<i>Sample</i>	<i>Mean (ng/ml)</i>	<i>SD (ng/ml)</i>	<i>CV (%)</i>
1	1541	1.68	10.9
2	1031	1.26	12.3

- **Spiking Recovery**

Serum samples were spiked with different amounts of rat Cystatin C, diluted with Dilution Buffer 100x and assayed.

<i>Sample</i>	<i>Observed (ng/ml)</i>	<i>Expected (ng/ml)</i>	<i>Recovery O/E (%)</i>
1	175.8	-	-
	297.4	300.8	98.9
	231.4	238.3	97.1
	213.7	207.1	103.2
2	189.4	-	-
	299.6	314.4	95.3
	251.9	251.9	100.0
	206.2	220.7	93.4

- **Linearity**

Serum samples were serially diluted with Dilution Buffer and assayed.

<i>Sample</i>	<i>Dilution</i>	<i>Observed (ng/ml)</i>	<i>Expected (ng/ml)</i>	<i>Recovery O/E (%)</i>
1	-	162.4	-	-
	2x	81.9	81.2	100.9
	4x	41.3	40.6	101.7
	8x	20.0	20.3	98.5
2	-	179.3	-	-
	2x	91.6	89.6	102.2
	4x	47.4	44.8	105.7
	8x	23.3	22.4	104.0

- **Stability of samples stored at 2-8C**

Samples should be stored at -20°C . However, no decline in concentration of Cystatin C was observed in serum samples after 14 days when stored at 2-8C. To avoid microbial contamination, samples were treated with ϵ -aminocaproic acid and sodium azide, resulting in the final concentration of 0.03% and 0.1%, respectively.

Sample	Incubation Temp. Period	Serum (ng/ml)
1	-20°C	102.0
	2-8 $^{\circ}\text{C}$, 7 day	105.9
	2-8 $^{\circ}\text{C}$, 14 day	113.5
2	-20°C	116.0
	2-8 $^{\circ}\text{C}$, 7 day	120.0
	2-8 $^{\circ}\text{C}$, 14 day	118.8
3	-20°C	115.4
	2-8 $^{\circ}\text{C}$, 7 day	100.4
	2-8 $^{\circ}\text{C}$, 14 day	120.7

- **Effect of Freezing/Thawing**

No decline was observed in concentration of rat Cystatin C in serum samples after repeated (3x) freeze/thaw cycles. However it is recommended to avoid unnecessary repeated freezing/thawing of the samples.

Sample	Number of f/t cycles	Serum (ng/ml)
1	1x	101.6
	3x	100.7
	5x	106.5
2	1x	106.1
	3x	92.2
	5x	95.7
3	1x	66.5
	3x	74.1
	5x	75.5

- **Reference ranges**

It is recommended that each laboratory include its own panel of control sample in the assay. Each laboratory should establish its own normal and pathological reference ranges for Cystatin C levels with the assay.

14. DEFINITION OF THE STANDARD

In this assay as the Standard the recombinant protein Cystatin C is used. This Cystatin C protein composed from 120 amino acid residues was produced in E.coli system. The apparent molecular weight is 14.93 kDa.

15. METHOD COMPARISON

Since no competitive commercial test is available, ALX-850-330 Rat Cystatin C ELISA has not been compared to any other immunoassay.

16. TROUBLESHOOTING AND FAQS

»» Weak signal in all wells

Possible explanations:

- Omission of a reagent or a step
- Improper preparation or storage of a reagent
- Assay performed before reagents were allowed to come to room temperature
- Improper wavelength when reading absorbance

»» High signal and background in all wells

Possible explanations:

- Improper or inadequate washing
- Overdeveloping; incubation time with Substrate Solution should be decreased before addition of Stop Solution
- Incubation temperature over 30°C

»» High coefficient of variation (CV)

Possible explanation:









- Improper or inadequate washing
- Improper mixing Standards, Quality Controls or samples

17. REFERENCES

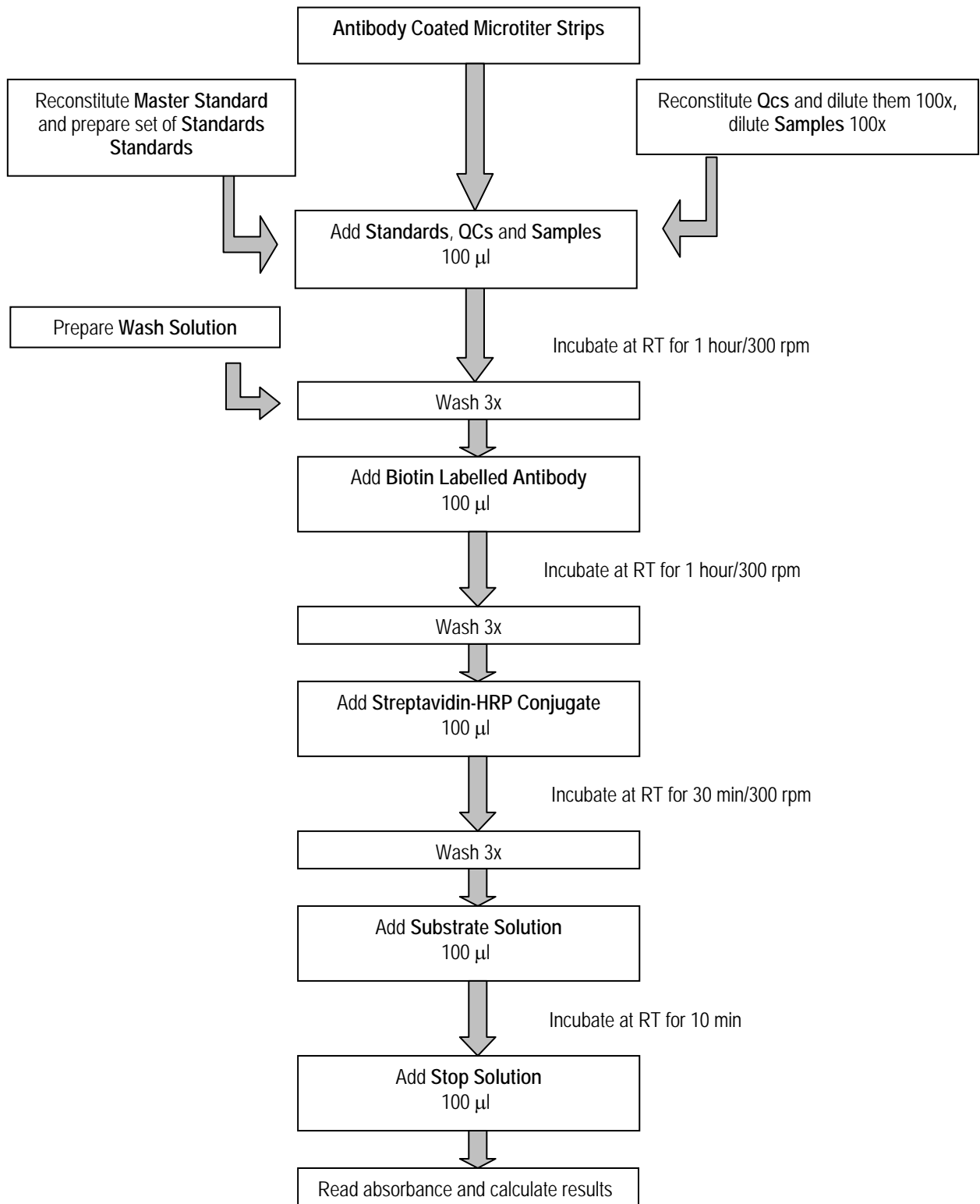
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18. EXPLANATION OF SYMBOLS

	Catalogue number
	Content
	Lot number
	See instructions for use
	Biological hazard
	Expiry date
	Storage conditions
	Identification of packaging materials

Assay Procedure Summary



1									
2									
3									
4									
5									
6									
7									
8									
9									
10									
11									
12									
	A	B	C	D	E	F	G	H	

NOTES



